



Article title: Neurogenesis and pattern separation: Time for a divorce

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Abstract

The generation of new neurons in the adult mammalian brain has led to numerous theories as to their functional significance. One of the most widely held views is that adult neurogenesis promotes pattern separation, a process by which overlapping patterns of neural activation are mapped to less overlapping representations. While a large body of evidence supports a role for neurogenesis in high interference memory tasks, it does not support the proposed function of neurogenesis in mediating pattern separation. Instead, the adult-generated neurons seem to generate highly overlapping and yet distinct distributed representations for similar events. One way in which these immature, highly plastic, hyperactive neurons may contribute to novel memory formation while avoiding interference is by virtue of their extremely sparse connectivity with incoming perforant path fibers. Another intriguing proposal, awaiting empirical confirmation, is that the young neurons' recruitment into memory formation is gated by a novelty / mismatch mechanism mediated by CA3 or hilar back-projections. Ongoing research into the intriguing link between neurogenesis, stress-related mood disorders and age-related neurodegeneration may lead to promising neurogenesis-based treatments for this wide range of clinical disorders.

Introduction

Over fifty years ago, Altman and Das made the remarkable discovery of adult neurogenesis in the rodent brain.^{1,2} This landmark finding called into question the long-held dogma that there are no new neurons created after birth. Not surprisingly, their discovery of neurogenesis met with intense scrutiny and scepticism. It took several decades of mounting evidence³⁻⁹ to convince the scientific community at large that there is indeed ongoing genesis of new neurons in the adult mammalian brain. Since the 1990's, an explosion of research into neurogenesis has taken place, from basic mechanisms to functional implications. Adult neurogenesis has now been reported in a wide range of species from rodents to primates, including macaques¹⁰ and humans¹¹⁻¹⁴.

Why was the phenomenon of adult neurogenesis so surprising? In short, the continuous addition of new neurons is the most extreme form of plasticity ever discovered in the adult mammalian brain. Consider what would happen if our entire brain remained neurogenic through the lifespan. Our memory circuits would be constantly re-wiring, forever erasing previously acquired knowledge and memories. Thus, under normal conditions, adult neurogenesis is restricted to very few mammalian brain regions, most notably, the dentate gyrus (DG) of the hippocampus and the olfactory bulb (OB).

In the DG of the adult rat, several thousand new granule cell neurons are generated every day¹⁵, representing less than one percent of the total cell population. In adult humans, levels are even lower; about 700 new DG neurons are generated in each hippocampus per day, comparable to levels found in mice¹⁴. Although this represents only a small percentage of the total DG cell population being generated and/or renewed each day, the unique properties of the young DG neurons make them well poised to contribute to behaviour. The young (4-6 week old) DG neurons are hyper-excitable and highly plastic¹⁶⁻¹⁹, so that they are recruited preferentially into novel memory traces^{16,18} relative to fully mature DG neurons. Similarly, in the olfactory bulb of rodents, thousands of new granule cell interneurons are generated each day²⁰. Relative to mature OB neurons, these newly generated young neurons exhibit an elevated and long-lasting responsiveness to new odours²¹. A key focus of ongoing research is how the brain is able to make use of these newly generated neurons to affect behaviour.

Adult Neurogenesis

Neurogenesis refers to the process by which new neurons are created. During development, this process occurs throughout the nervous system. In contrast, in the adult mammalian brain, genesis of new neurons has been reported across a wide range of mammalian species in only two regions: the dentate gyrus of the hippocampus and the subventricular zone-olfactory-bulb pathway. In the human brain, the preponderance of evidence indicates that there is very limited, if any, neurogenesis in the OB^{14,22}. Interneurons may also be generated in other regions of the adult mammalian brain, including the striatum and forebrain.²³⁻²⁵ Following a stroke or other brain injury, neurogenesis may occur in other areas of the mammalian brain as well^{26,27}. In contrast to mammals, in reptiles and birds, adult neurogenesis occurs in many regions of the adult brain²⁸⁻³⁰. Thus it appears that under normal conditions, as opposed to following a stroke or other brain trauma, greater brain complexity is associated with less widespread neurogenesis.

The process of neurogenesis begins with multi-potent neural progenitor cells or stem cells. These progenitor cells repeatedly undergo cell division and specialization into different cell types, some of which will be neurons. The entire process of neurogenesis includes this initial period of neural proliferation, followed by a period of pruning and maturation. Each of the young neurons either undergoes apoptosis (cell death) or survival and a prolonged period of maturation over the course of several weeks. A multitude of different factors promote one or both of these two components of neurogenesis: neuronal proliferation and survival.

Unique Properties of Immature Adult-Generated Neurons

The young, immature adult-generated neurons in the rodent dentate gyrus are affected very little by GABA inhibition, hence they fire more readily and are much more plastic than mature GCs, in spite of receiving fewer synaptic inputs^{16,18,19,31-33}. By 4 weeks of age they are preferentially recruited into memory circuits³⁴. As they mature further, they become more densely innervated by excitatory and inhibitory inputs; by age 6-8 weeks, they exert inhibitory control over other mature DG cells, and are themselves, in turn, regulated by feedback inhibition and fire very sparsely³³. Similarly, in the olfactory bulb (OB), newly generated granule cells are preferentially recruited to encode novel odours²¹. The OB granule cells participate in a unique form of lateral dendro-dendritic synaptic interaction with OB mitral cells (the principal relay neurons), transforming discrete spatial input patterns into complex distributed temporal patterns³⁵. This suggests that the OB granule cells, like the DG granule cells in the hippocampus, serve an important information-processing function. Thus, young adult-generated neurons in both the DG and OB are well positioned to be recruited selectively for new memory formation.

EVIDENCE OF A ROLE FOR NEUROGENESIS IN MEMORY

Does adult neurogenesis contribute significantly to behaviour? One way to address this question is to examine species that display natural variations in neurogenesis levels in the wild. For example, some species of birds exhibit seasonal changes in food-caching; their peak time of year for food caching and hence spatial learning, the autumn, coincides with a seasonal peak in neurogenesis levels³⁶. Rodent species who cache winter food in a single site have much lower neurogenesis levels than species that cache at multiple sites³⁷. Similarly, within-species geographical variations in food caching behaviour of red squirrels predict their neurogenesis levels³⁸. Thus, in both birds and rodents, neurogenesis levels co-vary with spatial learning and memory demands.

Memory impairments after neurogenesis knockdown

What types of memory is neurogenesis important for? The hippocampus is well established to be critical for learning and retrieving complex associative memories, including memory for sequences, contexts, spatial layouts, and episodes³⁹⁻⁴⁴. Many studies have investigated whether some or all of these established hippocampal-dependent memory functions may rely, more specifically, upon neurogenesis. The most common protocol is to apply a knock-down method that interferes with cell proliferation, wait several weeks so that any remaining young neurons have fully matured, and then test learning and memory. Knockdown methods include anti-mitotic drugs, low-dose focal brain irradiation, and permanent or temporary genetic manipulations⁴⁵⁻⁴⁸. More ecologically valid manipulations include binge ethanol consumption⁴⁹, high fat diets⁵⁰, chemotherapy drugs⁵¹, sleep deprivation⁵², stress^{53,54}, contextual fear learning⁵⁵ and manipulation of adrenal hormones^{7,56,57}. Further, there is an age-related decline in neurogenesis levels⁵⁸⁻⁶⁰.

Different methods for knocking down neurogenesis, across different species, have yielded somewhat contradictory results. Nonetheless, there is converging evidence from most (but not all) such studies

that neurogenesis knockdown disrupts performance on tasks that have a high interference component. One such task is contextual fear conditioning^{46,48,61,62}, where the animal has to associate a particular context with a subsequent aversive stimulus such as electric shock. Animals with intact neurogenesis levels exhibit a fear response specifically to the training context, whereas animals with low levels of neurogenesis will over-generalize their fear to different contexts, such as two different test boxes. Another potential source of interference in memory studies is a long time delay, whether the task involves a long delay between learning and the retention test, or the necessity to associate items across time. Importantly, animals with reduced neurogenesis exhibit memory deficits on long-term retention tests several weeks after learning in the Morris Water Maze (MWM)^{48,63,64}, associating stimuli across time delays (i.e. trace eyeblink conditioning)⁴⁵, and in delayed non-match to sample across long time delays⁴⁶. Another type of interference arises in tasks that require discriminating between similar items within an environment. Neurogenesis knockdown disrupts performance on tasks that require discriminating between similar spatial locations or objects⁶⁵ (sometimes called behavioural tests of pattern separation, but see next section), as well as discriminating similar contexts⁶⁶⁻⁶⁸. Finally, a memory task can have interference due to overlapping but conflicting information being learned at different points in time. Animals with reduced neurogenesis are also vulnerable to this type of interference, showing deficits on tasks that create retroactive interference between current and previously learned information⁶⁹, proactive interference between overlapping stimulus sets learned at different times⁷⁰, and interference due to reversal learning^{48,71-73} and extinction⁷⁴⁻⁷⁶. On the other hand, low interference versions of many of the above tasks are unaffected by a neurogenesis knockdown. Thus, neurogenesis is not required for initial acquisition in the MWM^{48,63,64} or simple fear conditioning^{46,61}. Similar findings on high interference olfactory tasks have been reported when OB neurogenesis is knocked down. Thus, in the OB, inhibition of postnatal neurogenesis impairs learning to discriminate highly overlapping odours, and long-term retention and reversal learning of olfactory associations, while leaving intact simple odour discrimination and odour associative learning.⁷⁷⁻⁸⁰ Moreover, post-training ablation of adult-generated neurons impairs previously learned odor-reward associations, contextual fear conditioning, and memory for spatial locations and visual discriminations in the MWM^{81,82}. Thus on tasks that may not require neurogenesis for acquisition, such as olfactory association learning, and spatial and non-spatial variants of the MWM, if the young neurons are recruited for memory formation, they will then be involved crucially in subsequent memory retrieval.

Positive effects of neurogenesis knock-down on memory

If knock-down of neurogenesis disrupts long-term memory, might it impart a *benefit* on tasks requiring short-term or working memory, when they are best performed by ignoring past memories? Saxe et al⁸³ created just such a task. In an 8-arm radial maze, rodents were required to remember which sequence of arms had been rewarded most recently, ignoring previously learned overlapping sequences. As predicted, rodents with reduced neurogenesis outperformed animals with intact neurogenesis levels on this task.

Memory enhancement after neurogenesis up-regulation

In addition to down-regulators, numerous extrinsic factors up-regulate neurogenesis. In the hippocampus, these include running⁸⁴, learning⁸⁵, environmental enrichment⁸⁶, dietary restriction⁸⁷ and dietary supplements that include anti-oxidant and anti-inflammatory factors⁸⁸⁻⁹². These up-

regulators can act on neurogenesis via dissociable mechanisms; running mainly affects proliferation whereas learning and enrichment increase neuronal survival⁹³. In the olfactory bulb, up-regulators of neurogenesis include exposure to olfactory enrichment and olfactory discrimination learning^{94–97} but not exercise⁹⁸.

In contrast to the effects of anti-neurogenic factors, as one would expect, pro-neurogenic factors *promote* performance on high interference memory tests, including contextual fear conditioning⁹⁹, discriminating similar spatial locations¹⁰⁰, and discriminating similar contexts^{67,101}. Moreover, pro-neurogenic factors such as running, environmental enrichment, and diet supplementation can mitigate the neurotoxic effects of alcohol exposure, stress, ageing, and irradiation on the brain and protect neurogenesis-dependent memory functions^{70,102–105}.

Memory impairment after neurogenesis up-regulation

Most studies that have investigated the functional effects of neurogenesis-upregulation have tested learning and memory after several weeks of an intervention such as exercise. Alternatively, one can up-regulate neurogenesis post-learning, to ask whether an increase in neurogenesis aids or interferes with memory retention. When adult rodents were exposed to wheel running after learning in a contextual fear conditioning task, this post-learning upregulation of neurogenesis was found to interfere with the previously acquired contextual fear conditioning response¹⁰⁶. One interpretation of these results is that a basic function of neurogenesis, when levels are elevated, is to promote memory clearance (see the “Memory clearance hypothesis” discussed in the next section)¹⁰⁶. As post-natal neurogenesis levels are highest in the infant brain, this also provides an explanation of infantile amnesia¹⁰⁶.

Converging evidence from human studies

Considering the importance of neurogenesis for memory in non-human animals, it is of great interest to know whether the same holds true in humans. In the absence of a non-invasive *in vivo* measure, all *direct* evidence of neurogenesis in the human adult brain comes from post-mortem assays^{11–14}. Several imaging methods show promise for assaying biological indicators of neurogenesis *in vivo*. In rodents, aerobic exercise up-regulates both neurogenesis and angiogenesis in the DG; these changes in angiogenesis can be detected using contrast-enhanced MRI of DG blood volume and correlate with increased neurogenesis, while in humans, the same exercise-induced increase in DG blood volume is observed after several weeks of exercise.¹⁰⁷ Other promising methods for imaging neurogenesis indicators include MR spectroscopy¹⁰⁸ and PET¹⁰⁹, although current methods lack sufficient specificity¹¹⁰.

Several investigators have developed human analogues of neurogenesis-dependent cognitive tests used in rodents. Extrinsic up- and down-regulators of neurogenesis in rodents should have similar impact on human performance. Consistent with this prediction, humans with symptoms of a first episode of depression were impaired on the CANTAB delayed match to sample task¹¹¹, which tests delayed recognition memory for images of abstract, complex objects amongst a set of highly overlapping lures. Elevated stress and depression scores also predict impairments on the Mnemonic Similarity Task (MST) and variants^{112,113}, which test memory for images of every day objects versus highly similar lures. Conversely, aerobic response to several weeks of exercise correlates with changes in performance in the MST¹¹³. Finally, those with elevated stress, depression and binge

alcohol scores are more impaired on tests of memory for overlapping spatial locations¹¹⁴. Thus, converging evidence across species suggests that hippocampal neurogenesis plays a similar role in human memory to that in rodents.

Common factors across neurogenesis-dependent tasks: Overcoming interference

The evidence reviewed above indicates that neurogenesis is required for many different memory tasks. These include distinguishing recently studied items from spatially or visually overlapping lures, learning distinct representations for items encountered in similar contexts, memory for items across long time delays, and extinguishing or reversing previously responses. The commonality across this wide range of seemingly disparate neurogenesis-dependent tasks is that they require memory representations that are robust against many different types of interference. In the next section, we consider alternative theoretical perspectives on the function of neurogenesis that attempt to account for these and other findings.

THEORETICAL PERSPECTIVES

Pattern separation

One of the most widely proposed functions of neurogenesis is to promote pattern separation. The term pattern separation was coined by computational modellers to refer to a type of neural coding, whereby overlapping input patterns are coded as less overlapping output codes. One way to achieve a less overlapping output code is via sparse coding, as illustrated in Figure 1a. Given that pattern separation is a characteristic of the neural code, pattern separation can only be verified by recording neural activation patterns. Nonetheless, many researchers use the term “behavioural pattern separation” to refer to almost any behavioural task that has a high interference component, assuming that a behavioural assay correlates with the underlying, hypothesized neural code.

Evidence of pattern separation in the dentate gyrus

While there is some empirical support for the role of the DG in pattern separation (in its original sense), the role of neurogenesis in this process remains controversial. Computational models of memory with sparse coding confirm that sparse coding leads to greater pattern separation, and that pattern separation is an effective mechanism for mitigating memory interference^{115–118}. Given the extremely sparse firing rates of neurons in the DG¹¹⁹, many modellers have adopted the assumption that pattern separation is a fundamental computational function of the DG, while pattern completion (cued memory retrieval) is a function of the CA3 region^{115–118,120–122}. This caricature of a hippocampus that performs pattern separation in the DG and pattern completion in CA3 has become pervasive in the literature. However, like any caricature, it captures some key features, while ignoring many important details. Findings from electrophysiological and immediate early gene activation studies confirm that the DG robustly differentiates distinct contexts and environments, even based on very subtle features such as task demands, by recruiting different subsets of granule cells^{119,123–126}, consistent with sparse coding and pattern separation in the DG. Evidence from human fMRI studies lends further support to this notion¹²⁷. Importantly, however, some of these activation studies also paint a more nuanced picture, as a large subset (about 30%) of granule cells are jointly recruited when an animal is exposed to two different contexts or environments, or even

the same environment under different task demands^{123,125,128–130}, a finding that is inconsistent with sparse coding and pattern separation. One possible explanation for these findings is that the subset of neurons activated across multiple contexts is the hyperactive, immature neuron population.

Does neurogenesis contribute to pattern separation?

The extensive evidence of a role for hippocampal neurogenesis in high interference behavioural tasks (which are often referred to as “behavioural pattern separation” tasks, a highly problematic term), led to the suggestion that neurogenesis could be directly responsible for pattern separation¹³¹. However, such a direct relation seems unlikely given that immature (4-6 week old) neurons are hyperactive rather than firing sparsely. Computational modellers have established that in neural models that have higher activity levels, there is a greater probability of overlap between neural codes for different memories, whereas in neural models that employ sparse codes, there is greater pattern separation (i.e. less pattern overlap).¹³² Indeed, a recent model of the dentate gyrus demonstrates that the addition of highly active young neurons decreases sparse coding, decreases pattern separation, and yet improves memory performance.¹³³

An updated view is that the young neurons may increase sparse coding, leading to increased pattern separation, in an indirect manner, by recruiting greater feedback inhibition onto mature granule cells^{134–136}. The increased recruitment of feedback inhibition over mature granule cells is also consistent with the hypothesis of a circuit-level homeostatic mechanism that regulates overall activity within the DG¹³⁷. Thus, high neurogenesis levels, translating into high activity levels in the immature granule cell population, require a compensatory lowering of activity levels in the mature cell population in order to balance overall activity in the DG.

Pattern integration and the memory resolution hypothesis

Given that the behaviour of the young immature neurons is inconsistent with the pattern separation hypothesis, how else might these neurons contribute to memory encoding? A very different idea is that the immature neurons, by firing continuously over time, may function as pattern integrators rather than pattern separators.^{138–140} Thus, the immature neurons could provide the representation of temporal context that binds together elements of an episode. In support of this idea, electrophysiological recordings reveal distinct pools of DG neurons activated in different contexts that are well separated in time; either decreasing this temporal separation or knocking down neurogenesis attenuates their contextual selectivity, such that many of the young neurons fire in distinct but similar contexts.¹⁴¹

A more recent advancement on the above idea is the memory resolution hypothesis, which proposes complementary roles for the immature and mature neurons. The young immature neurons are coarsely tuned to a wide range of features, allowing them to better represent new information, while the mature neurons encode information at a high resolution, minimizing overlap between memory representations^{142–144}. This account attributes the pattern separation function to the mature granule cells (enhanced by feedback inhibition from immature neurons), and the pattern integration function to immature neurons. However, this account has trouble explaining how highly similar items could be encoded within the same context. Given that mature granule cells are not very plastic relative to immature neurons, and tightly tuned to contexts that were learned,

presumably, when they were at an immature stage, how could they be recruited to differentiate novel, similar features?

The Memory clearance hypothesis

Rather than promoting memory formation, a rather different role has been proposed for newly generated neurons in memory clearance.^{137,145–149} This could be a means by which the brain clears out older, more remote memories in favour of novel memory encoding. Such a role in memory clearance is not necessarily incompatible with a dual role for neurogenesis in supporting novel memory formation. There are several lines of empirical support for the memory clearance hypothesis. For example, mice with elevated levels of neurogenesis exhibit compromised memory stability.¹⁵⁰ Importantly, the memory clearance hypothesis also provides a compelling explanation of infantile amnesia¹⁴⁹, as neurogenesis levels in mammalian hippocampi are highest at birth, and decline with age. On the other hand, the memory clearance hypothesis is difficult to reconcile with direct evidence from rodents,⁶⁹ and indirect evidence from humans,¹⁵¹ that higher neurogenesis levels protect remote memories from retroactive interference with newly acquired information. Given the support for both of these theories, interference reduction versus memory clearance, an important avenue for further research is to discover how these apparently contradictory lines of evidence can be reconciled.

CONTROVERSIES

Could young neurons decrease pattern separation while decreasing interference?

One issue that is difficult to explain for the theories described so far is that young neurons seem to *decrease* pattern separation, and yet they are crucial for mitigating interference. The evidence reviewed above indicates that the young neurons are required for a wide range of high interference tasks, whether the interference arises between similar stimuli in the same context, or similar events in different contexts separated in time. Importantly, while neurogenesis levels decline with age, pattern separation increases with age, as measured by activation patterns in the DG using immediate early gene labelling¹²⁸. Moreover, animals' ability to discriminate two contexts is positively predicted by the overlap: overlapping representations are associated with improved discrimination¹²⁸.

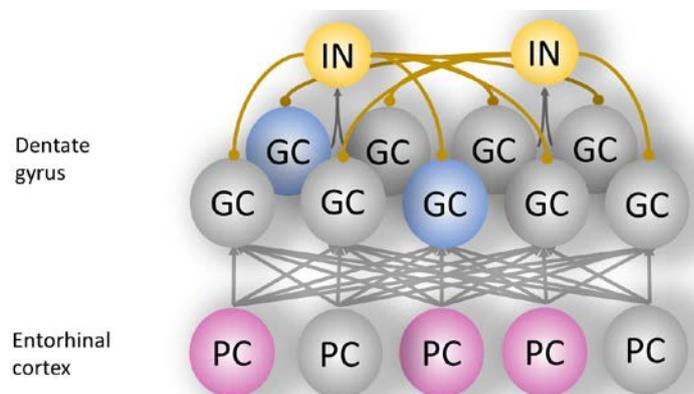
Sparse connectivity decreases interference

Sparse coding / pattern separation is not the only strategy for overcoming interference. In fact, under conditions of very high plasticity, it does not solve the problem. Computer simulations of "Competitive Learning" neural networks^{152–154}, with ultra-sparse coding enforced via a winner-take-all activation function, require very slow adjustment to the synaptic weights, and many iterations through a set of training patterns (as opposed to one-shot learning), for proper functioning. Otherwise, they suffer from interference with previous learning, illustrating the stability-plasticity tradeoff¹⁵⁴.

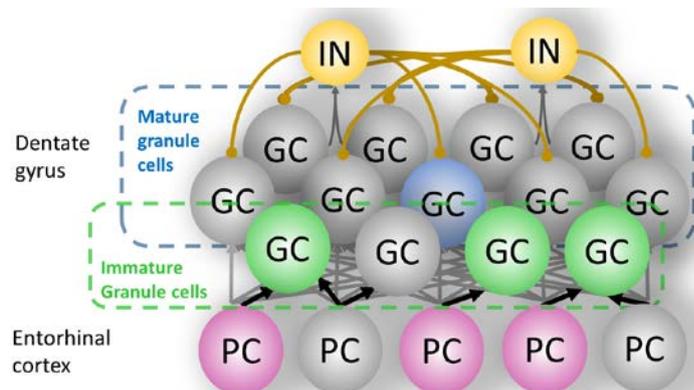
One strategy for overcoming interference, while maintaining high levels of plasticity, is to have sparse connectivity. This seems to be the strategy used by the young immature neurons. They are

very sparsely connected, receiving relatively few inputs from the entorhinal cortex, and few if any inputs from lateral neighbors or descending inputs from CA3 neurons; if they survive and mature, they undergo experience-dependent synaptic remodelling, becoming increasingly innervated by cortical and hippocampal inputs¹⁵⁵. As illustrated in Figure 1b) and c), a pool of young, sparsely connected, immature granule cells can respond to two different overlapping stimuli with overlapping and yet distinct patterns of activation. The sparse connectivity of the immature granule cells increases the tendency for each neuron to respond to a different subset of input features. Recent empirical evidence supports the notion that the low synaptic connectivity of immature neurons prevents them from firing broadly to a wide range of stimuli.¹⁵⁶ Computer simulations of a model with these characteristics lend further support to the proposal that immature neurons with high plasticity and sparse connectivity are able to overcome interference, while simultaneously *decreasing* pattern separation¹³³.

A



B



C

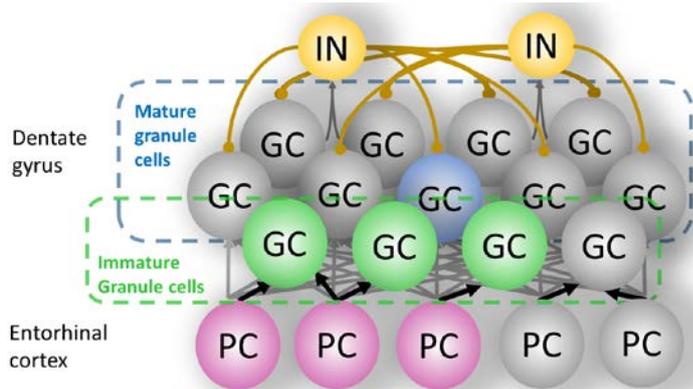


Figure 1. Schematic models of the hippocampus, using sparse coding (A) versus neurogenesis and sparse connectivity (B, C) to overcome interference. Model input is a distributed pattern of activation across the entorhinal cortex pyramidal cells (PC; activated cells shown in pink). This input generates a sparse pattern of activation across the mature dentate granule cells (GCs; active cells shown in blue). PCs are densely interconnected to mature GCs, which are themselves interconnected via inhibitory interneurons (IN, shown in yellow). In B) and C), the model with neurogenesis also includes immature GCs (active cells shown in green); entorhinal PCs are sparsely connected to the immature GCs. The model shown in B) and C) is presented with two different input patterns and in response, generates identical sparse activation patterns in mature GCs, and overlapping but distinct distributed activation in immature GCs. While the two input patterns overlap by 40%, the two patterns of activation in the immature GC overlap by 50%, hence a decrease in pattern separation. The model is nonetheless capable of maintaining distinct neural codes for the two similar inputs, in spite of high plasticity in the immature cell population, due to the sparse connectivity of the immature GCs.

Another potential strategy for overcoming interference, while maintaining high levels of plasticity, is to use top-down expectations from descending pathways to compute a novelty / mismatch signal, and have the mismatch signal drive recruitment of new neurons for encoding new memories. Such a mechanism was proposed several decades ago in a model of classification learning known as Adaptive Resonance Theory^{154,157}. Computational models of mismatch detection within the DG-CA3 circuit^{122,158}, together with feedback projections from the CA3 to DG, could explain how novelty/ mismatch signals might drive DG circuit dynamics, inhibitory feedback and plasticity. Such a mechanism could allow the circuit to operate in two distinct modes, memory storage versus recall.

The fate of old neurons and old memories

A major unresolved issue is what happens to adult-born neurons as they continue to age. Evidence suggests that adult-generated neurons recruited during memory acquisition are also recruited for subsequent memory retrieval¹⁵⁹. However, behaviourally, it is virtually impossible to distinguish memory formation from memory retrieval, as memories may be modified whenever they are retrieved. Indeed, the recruitment of young immature neurons has been implicated in this process of memory reconsolidation¹⁶⁰.

Evidence from neurogenesis knockdown studies suggests an important role for neurogenesis in long-term retention and remote memory^{48,63,64}, also supported by correlative evidence in humans¹⁵¹. On the other hand, it has been found that as the mature granule cells age, they become less and less active, and eventually may retire into silence¹³⁰. It would be surprising if this was the fate of all adult-generated neurons, but this is an unresolved issue.

The neurogenic theory of depression

The intriguing link between stress, depression and neurogenesis led to the neurogenic theory of depression¹⁶¹. According to this view, reduced neurogenesis causes depression, and restoration of neurogenesis leads to the recovery from depression. In support of this view, stress is well established to reduce neurogenesis¹⁶², it is widely believed to play a major role in causing depression, and is the basis of all animal models of depression. Further, many anti-depressant factors, including SSR's, ECT, aerobic exercise, and successful stress coping, up-regulate neurogenesis in animal models^{84,163-168}. What remains a matter of debate is whether neurogenesis plays a causal role in either the pathogenesis of depression or in its recovery. Suppressed neurogenesis by itself does not cause a depressive or anxious phenotype, leading to the suggestion that it is not directly involved in mood regulation, but instead modulates emotional responding via its role in mnemonic processing¹⁶⁹. On the other hand, in rodent models, neurogenesis knockdown increases the HPA axis response¹⁷⁰ and predisposes the animal to be more sensitive to the effects of stress¹⁷¹, while neurogenesis knockdown blocks the anti-depressant effects of SSRIs^{172,173}.

Conclusion

While empirical studies point to a role for neurogenesis in reducing interference between similar events in memory, the mechanism by which this interference reduction is achieved is still under debate. Earlier theories postulated a role for the young neurons in pattern separation, a mechanism by which similar neural activation patterns are encoded as very sparse, less overlapping representations. However, the immature neurons do not behave in a manner consistent with the proposed pattern separation function. The evidence reviewed above indicates that these neurons fire at low thresholds, generating highly overlapping neural codes for similar events, and yet, they are crucial for distinguishing similar events or contexts. An alternative view is that the young neurons generate distributed codes across similar contexts that are overlapping, but nonetheless distinct, by using sparse connectivity. Their sparse perforant path afferent connections cause the immature neurons to maintain some degree of selectivity in spite of very high plasticity levels. Additionally, top-down mismatch signals from the CA3 region could play a role in regulating activity and plasticity levels in the DG, and gating the operation of the circuit between storage and recall modes. For such a scheme to work, the top-down mismatch signal would have to gate the recruitment of immature DG neurons for novel memory encoding. While this general scheme is intriguing, it remains to be worked out how such a function could operate within the DG / CA3 circuit, and for such a model to be validated empirically.

An important future application of neurogenesis research is in the treatment of stress-related neuropsychiatric disorders. While a causal link between neurogenesis and depression has yet to be established, an emerging view is that animals with reduced neurogenesis have an impairment at

encoding and recognizing contexts, resulting in an over-generalization of fear and an increased vulnerability to mood disorders.¹⁷⁴ Conversely, rescue of neurogenesis may support the normal hippocampal role in exerting contextual modulation over neural circuits subserving stress, emotion and other responses.^{175,176}

Neurogenesis research also has important implications for treating age-related neuropathology. Factors associated with age-related neurodegeneration and dementia, including a dysregulated HPA axis, chronic inflammation, and microglial activation, also impair neurogenesis^{60,91,177–179}. Thus, a promising target for interventions in this wide range of disorders is to up-regulate neurogenesis levels and associated neurotrophic factors. Increasing neurogenesis and neurotrophic factor levels via exercise, diet, and environmental enrichment may impart neuroprotection against stress, ageing and dementia^{104,180–184}. Also, there may be important interactions amongst these factors in promoting optimal learning^{185,186} and a healthy brain^{104,187}.

References

1. Altman, J. & Das, G. D. Autoradiographic and histological evidence of postnatal hippocampal neurogenesis in rats. *J. Comp. Neurol.* **124**, 319–335 (1965).
2. Altman, J. Are New Neurons Formed in the Brains of Adult Mammals? *Science (80-.)*. **135**, 1127–1128 (1962).
3. Kaplan, M. & Hinds, J. Neurogenesis in the adult rat: electron microscopic analysis of light radioautographs. *Science (80-.)*. **197**, 1092–1094 (1977).
4. Kaplan, M. S. & Bell, D. H. Mitotic neuroblasts in the 9-day-old and 11-month-old rodent hippocampus. *J. Neurosci.* **4**, 1429–1441 (1984).
5. Wyss, J. M. & Sripanidkulchai, B. The development of Ammon's horn and the fascia dentata in the cat: a [3H]thymidine analysis. *Brain Res.* **350**, 185–198 (1985).
6. Altman, J. & Bayer, S. A. Migration and distribution of two populations of hippocampal granule cell precursors during the perinatal and postnatal periods. *J. Comp. Neurol.* **301**, 365–381 (1990).
7. Gould, E., Woolley, C. S., Cameron, H. A., Daniels, D. C. & McEwen, B. S. Adrenal steroids regulate postnatal development of the rat dentate gyrus: II. Effects of glucocorticoids and mineralocorticoids on cell birth. *J. Comp. Neurol.* **313**, 486–493 (1991).
8. Cameron, H. A., Woolley, C. S., McEwen, B. S. & Gould, E. Differentiation of newly born neurons and glia in the dentate gyrus of the adult rat. *Neuroscience* **56**, 337–344 (1993).
9. Lois, C. & Alvarez-Buylla, A. Long-distance neuronal migration in the adult mammalian brain. *Science (80-.)*. **264**, 1145–1148 (1994).
10. Kornack, D. R. & Rakic, P. Continuation of neurogenesis in the hippocampus of the adult macaque monkey. *Proc. Natl. Acad. Sci.* **96**, 5768–5773 (1999).
11. Eriksson, P. S. *et al.* Neurogenesis in the adult human hippocampus. *Nat. Med.* **4**, 1313–1317 (1998).
12. Bédard, A. & Parent, A. Evidence of newly generated neurons in the human olfactory bulb. *Dev. Brain Res.* **151**, 159–168 (2004).

13. Knoth, R. *et al.* Murine features of neurogenesis in the human hippocampus across the lifespan from 0 to 100 years. *PLoS One* **5**, e8809 (2010).
14. Spalding, K. L. *et al.* Dynamics of hippocampal neurogenesis in adult humans. *Cell* **153**, 1219–1227 (2013).
15. Cameron, H. A. & McKay, R. D. G. Adult Neurogenesis Produces a Large Pool of New Granule Cells in the Dentate Gyrus. *J. Comp. Neurol.* **435**, 406–417 (2001).
16. Wang, S., Scott, B. W. & Wojtowicz, J. M. Heterogenous properties of dentate granule neurons in the adult rat. *J. Neurobiol.* **42**, 248–257 (2000).
17. Snyder, J. S., Kee, N. & Wojtowicz, J. M. Effects of adult neurogenesis on synaptic plasticity in the rat dentate gyrus. *J Neurophysiol* **85**, 2423–2431 (2001).
18. Schmidt-Hieber, C., Jonas, P. & Bischofberger, J. Enhanced synaptic plasticity in newly generated granule cells of the adult hippocampus. *Nature* **429**, 184–187 (2004).
19. Marín-Burgin, A., Mongiat, L. A., Pardi, M. B. & Schinder, A. F. Unique processing during a period of high excitation/inhibition balance in adult-born neurons. *Science* (80-.). **335**, 1238–1242 (2012).
20. Alonso, M. *et al.* Activation of adult-born neurons facilitates learning and memory. *Nat. Neurosci.* **15**, 897–904 (2012).
21. Magavi, S. S. P., Mitchell, B. D., Szentirmai, O., Carter, B. S. & Macklis, J. D. Adult-Born and Preexisting Olfactory Granule Neurons Undergo Distinct Experience-Dependent Modifications of their Olfactory Responses In Vivo. *J. Neurosci.* **25**, 10729–10739 (2005).
22. Wang, C. *et al.* Identification and characterization of neuroblasts in the subventricular zone and rostral migratory stream of the adult human brain. *Cell Res.* **21**, 1534–1550 (2011).
23. Bédard, A., Cossette, M., Lévesque, M. & Parent, A. Proliferating cells can differentiate into neurons in the striatum of normal adult monkey. *Neurosci. Lett.* **328**, 213–216 (2002).
24. Dayer, A. G., Cleaver, K. M., Abouantoun, T. & Cameron, H. A. New GABAergic interneurons in the adult neocortex and striatum are generated from different precursors. *J. Cell Biol.* **168**, 415–427 (2005).
25. Ernst, A. *et al.* Neurogenesis in the striatum of the adult human brain. *Cell* **156**, 1072–1083 (2014).
26. Parent, J. M., Vexler, Z. S., Gong, C., Derugin, N. & Ferriero, D. M. Rat forebrain neurogenesis and striatal neuron replacement after focal stroke. *Ann. Neurol.* **52**, 802–813 (2002).
27. Arvidsson, A., Collin, T., Kirik, D., Kokaia, Z. & Lindvall, O. Neuronal replacement from endogenous precursors in the adult brain after stroke. *Nat. Med.* **8**, 963–970 (2002).
28. Font, E., Desfilis, E., Pérez-Cañellas, M. M. & García-Verdugo, J. M. Neurogenesis and neuronal regeneration in the adult reptilian brain. in *Brain. Behav. Evol.* **58**, 276–295 (2001).
29. Barnea, A. & Nottenbhom, F. Seasonal recruitment of hippocampal neurons in adult free-ranging black-capped chickadees. *PNAS* **91**, 11217–11221 (1994).
30. Barnea, A. & Pravosudov, V. Birds as a model to study adult neurogenesis: Bridging

- evolutionary, comparative and neuroethological approaches. *Eur. J. Neurosci.* **34**, 884–907 (2011).
31. Mongiat, L. A., Espósito, M. S., Lombardi, G. & Schinder, A. F. Reliable Activation of Immature Neurons in the Adult Hippocampus. *PLoS One* **4**, e5320 (2009).
 32. Li, Y., Aimone, J. B., Xu, X., Callaway, E. M. & Gage, F. H. Development of GABAergic inputs controls the contribution of maturing neurons to the adult hippocampal network. *Proc. Natl. Acad. Sci.* **109**, 4290–4295 (2012).
 33. Temprana, S. G. *et al.* Delayed Coupling to Feedback Inhibition during a Critical Period for the Integration of Adult-Born Granule Cells. *Neuron* **85**, 116–130 (2015).
 34. Kee, N., Teixeira, C. M., Wang, A. H. & Frankland, P. W. Preferential incorporation of adult-generated granule cells into spatial memory networks in the dentate gyrus. *Nat. Neurosci.* **10**, 355–362 (2007).
 35. Gheusi, G. & Lledo, P. Control of early events in olfactory processing by adult neurogenesis. *Chem. Senses* **32**, 397–409 (2007).
 36. Sherry, D. F. & Hoshoooley, J. S. Seasonal hippocampal plasticity in food-storing birds. *Philos. Trans. R. Soc. B Biol. Sci.* **365**, 933–943 (2010).
 37. Barker, J. M., Wojtowicz, J. M. & Boonstra, R. Where's my dinner? Adult neurogenesis in free-living food-storing rodents. *Genes, Brain Behav.* **4**, 89–98 (2004).
 38. Johnson, K. M., Boonstra, R. & Wojtowicz, J. M. Hippocampal neurogenesis in food-storing red squirrels: the impact of age and spatial behavior. *Genes, Brain Behav.* no–no (2010). doi:10.1111/j.1601-183X.2010.00589.x
 39. Tulving, E. & Markowitsch, H. J. Episodic and declarative memory: role of the hippocampus. *Hippocampus* **8**, 198–204 (1998).
 40. Chun, M. M. & Phelps, E. a. Memory deficits for implicit contextual information in amnesic subjects with hippocampal damage. *Nat. Neurosci.* **2**, 844–7 (1999).
 41. Fortin, N. J., Agster, K. L. & Eichenbaum, H. B. Critical role of the hippocampus in memory for sequences of events. *Nat. Neurosci.* **5**, 458–62 (2002).
 42. King, J. A., Burgess, N., Hartley, T., Vargha-Khadem, F. & O'Keefe, J. *Human hippocampus and viewpoint dependence in spatial memory.* *Hippocampus* **12**, 811–820 (2002).
 43. Bohbot, V. D., Iaria, G. & Petrides, M. Hippocampal Function and Spatial Memory: Evidence From Functional Neuroimaging in Healthy Participants and Performance of Patients With Medial Temporal Lobe Resections. *Neuropsychology* **18**, 418–425 (2004).
 44. Smith, D. M. & Mizumori, S. J. Y. Hippocampal place cells, context, and episodic memory. *Hippocampus* **16**, 716–29 (2006).
 45. Shors, T. J. *et al.* Neurogenesis in the adult is involved in the formation of trace memories. *Nature* **410**, 372–376 (2001).
 46. Winocur, G., Wojtowicz, J. M., Sekeres, M., Snyder, J. S. & Wang, S. Inhibition of neurogenesis interferes with hippocampus-dependent memory function. *Hippocampus* **16**, 296–304 (2006).
 47. Jessberger, S. *et al.* Dentate gyrus-specific knockdown of adult neurogenesis impairs spatial and object recognition memory in adult rats. *Learn. Mem.* **16**, 147–154 (2009).
 48. Pan, Y.-W., Chan, G. C. K., Kuo, C. T., Storm, D. R. & Xia, Z. Inhibition of Adult

- Neurogenesis by Inducible and Targeted Deletion of ERK5 Mitogen-Activated Protein Kinase Specifically in Adult Neurogenic Regions Impairs Contextual Fear Extinction and Remote Fear Memory. *J. Neurosci.* **32**, 6444–6455 (2012).
49. Nixon, K. & Crews, F. T. Binge ethanol exposure decreases neurogenesis in adult rat hippocampus. *J Neurochem* **83**, 1087–1093 (2002).
 50. Lindqvist, A. *et al.* High-fat diet impairs hippocampal neurogenesis in male rats. *Eur. J. Neurol.* **13**, 1385–1388 (2006).
 51. Christie, L. A. *et al.* Impaired cognitive function and hippocampal neurogenesis following cancer chemotherapy. *Clin. Cancer Res.* **18**, 1954–1965 (2012).
 52. Guzman-Marin, R. *et al.* Sleep deprivation suppresses neurogenesis in the adult hippocampus of rats. *Eur. J. Neurosci.* **22**, 2111–2116 (2005).
 53. Gould, E., McEwen, B. S., Tanapat, P., Galea, L. A. M. & Fuchs, E. Neurogenesis in the dentate gyrus of the adult tree shrew is regulated by psychosocial stress and MDA receptor activation. *J. Neurosci.* **17**, 2492–2498 (1997).
 54. Warner-Schmidt, J. L. & Duman, R. S. Hippocampal neurogenesis: Opposing effects of stress and antidepressant treatment. *Hippocampus* **16**, 239–249 (2006).
 55. Pham, K., McEwen, B. S., Ledoux, J. E. & Nader, K. Fear learning transiently impairs hippocampal cell proliferation. *Neuroscience* **130**, 17–24 (2005).
 56. Gould, E., Cameron, H. A., Daniels, D. C., Woolley, C. S. & McEwen, B. S. Adrenal Hormones Suppress Cell Division in the Adult Rat Dentate Gyrus. *J. Neurosci.* **12**, 3642–3650 (1992).
 57. Cameron, H. A. & Gould, E. Adult neurogenesis is regulated by adrenal steroids in the dentate gyrus. *Neuroscience* **61**, 203–209 (1994).
 58. Kuhn, H. G., Dickinson-Anson, H. & Gage, F. H. Neurogenesis in the dentate gyrus of the adult rat: Age-related decrease of neuronal progenitor proliferation. *J. Neurosci.* **16**, 2027–2033 (1996).
 59. McDonald, H. Y. & Wojtowicz, J. M. Dynamics of neurogenesis in the dentate gyrus of adult rats. *Neurosci Lett* **385**, 70–75 (2005).
 60. Montaron, M. F. *et al.* Lifelong corticosterone level determines age-related decline in neurogenesis and memory. *Neurobiol. Aging* **27**, 645–654 (2006).
 61. Saxe, M. D. *et al.* Ablation of hippocampal neurogenesis impairs contextual fear conditioning and synaptic plasticity in the dentate gyrus. *Proc Natl Acad Sci U S A* **103**, 17501–17506 (2006).
 62. Park, H. *et al.* Mice lacking the PSD-95–interacting E3 ligase, Dorfin/Rnf19a, display reduced adult neurogenesis, enhanced long-term potentiation, and impaired contextual fear conditioning. *Sci. Rep.* **5**, 16410 (2015).
 63. Snyder, J., Hong, N., McDonald, R. & Wojtowicz, J. A role for adult neurogenesis in spatial long-term memory. *Neuroscience* **130**, 843–852 (2005).
 64. Ben Abdallah, N. M. B. *et al.* Impaired long-term memory retention: Common denominator for acutely or genetically reduced hippocampal neurogenesis in adult mice. *Behav. Brain Res.* **252**, 275–286 (2013).
 65. Clelland, C. D. *et al.* A functional role for adult hippocampal neurogenesis in spatial pattern separation. *Science* **325**, 210–3 (2009).
 66. Kheirbek, M. A., Tannenholz, L. & Hen, R. NR2B-Dependent Plasticity of Adult-Born

- Granule Cells is Necessary for Context Discrimination. *J. Neurosci.* **32**, 8696–8702 (2012).
67. Nakashiba, T. *et al.* Young dentate granule cells mediate pattern separation, whereas old granule cells facilitate pattern completion. *Cell* **149**, 188–201 (2012).
 68. Niibori, Y. *et al.* Suppression of adult neurogenesis impairs population coding of similar contexts in hippocampal CA3 region. *Nat. Commun.* **3**, 1253 (2012).
 69. Luu, P. *et al.* The role of adult hippocampal neurogenesis in reducing interference. *Behav. Neurosci.* **126**, 381–91 (2012).
 70. Winocur, G., Becker, S., Luu, P., Rosenzweig, S. & Wojtowicz, J. M. Adult hippocampal neurogenesis and memory interference. *Behav. Brain Res.* **227**, 464–9 (2012).
 71. Burghardt, N. S., Park, E. H., Hen, R. & Fenton, A. A. Adult-born hippocampal neurons promote cognitive flexibility in mice. *Hippocampus* **22**, 1795–1808 (2012).
 72. Garthe, A., Behr, J. & Kempermann, G. Adult-generated hippocampal neurons allow the flexible use of spatially precise learning strategies. *PLoS One* **4**, (2009).
 73. Kalm, M., Karlsson, N., Nilsson, M. K. L. & Blomgren, K. Loss of hippocampal neurogenesis, increased novelty-induced activity, decreased home cage activity, and impaired reversal learning one year after irradiation of the young mouse brain. *Exp. Neurol.* **247**, 402–409 (2013).
 74. Deng, W., Saxe, M. D., Gallina, I. S. & Gage, F. H. Adult-born hippocampal dentate granule cells undergoing maturation modulate learning and memory in the brain. *J. Neurosci.* **29**, 13532–42 (2009).
 75. Noonan, M. A., Bulin, S. E., Fuller, D. C. & Eisch, A. J. Reduction of Adult Hippocampal Neurogenesis Confers Vulnerability in an Animal Model of Cocaine Addiction. *J. Neurosci.* **30**, 304–315 (2010).
 76. Cleva, R. M., Wischerath, K. C. & Olive, M. F. Extinction Learning and Adult Neurogenesis. *Neuropsychopharmacology* **36**, 360–361 (2011).
 77. Moreno, M. M. *et al.* Olfactory perceptual learning requires adult neurogenesis. *Proc. Natl. Acad. Sci. U. S. A.* **106**, 17980–17985 (2009).
 78. Sultan, S. *et al.* Learning-dependent neurogenesis in the olfactory bulb determines long-term olfactory memory. *FASEB J.* **24**, 2355–2363 (2010).
 79. Mak, G. K. & Weiss, S. Paternal recognition of adult offspring mediated by newly generated CNS neurons. *Nat. Neurosci.* **13**, 753–758 (2010).
 80. Sakamoto, M. *et al.* Continuous Postnatal Neurogenesis Contributes to Formation of the Olfactory Bulb Neural Circuits and Flexible Olfactory Associative Learning. *J. Neurosci.* **34**, 5788–5799 (2014).
 81. Arruda-Carvalho, M., Sakaguchi, M., Akers, K. G., Josselyn, S. A. & Frankland, P. W. Posttraining Ablation of Adult-Generated Neurons Degrades Previously Acquired Memories. *J. Neurosci.* **31**, 15113–15127 (2011).
 82. Arruda-Carvalho, M. *et al.* Posttraining Ablation of Adult-Generated Olfactory Granule Cells Degrades Odor-Reward Memories. *J. Neurosci.* **34**, 15793–15803 (2014).
 83. Saxe, M. D. *et al.* Paradoxical influence of hippocampal neurogenesis on working memory. *Proc. Natl. Acad. Sci. U. S. A.* **104**, 4642–6 (2007).

84. van Praag, H., Kempermann, G. & Gage, F. H. Running increases cell proliferation and neurogenesis in the adult mouse dentate gyrus. *Nat. Neurosci.* **2**, 266–270 (1999).
85. Gould, E., Beylin, A., Tanapat, P., Reeves, A. & Shors, T. J. Learning enhances adult neurogenesis in the hippocampal formation. *Nat. Neurosci.* **2**, 260–265 (1999).
86. Kempermann, G., Kuhn, H. G. & Gage, F. H. More hippocampal neurons in adult mice living in an enriched environment. *Nature* **386**, 493–495 (1997).
87. Lee, J., Seroogy, K. B. & Mattson, M. P. Dietary restriction enhances neurotrophin expression and neurogenesis in the hippocampus of adult mice. *J. Neurochem.* **80**, 539–547 (2002).
88. Casadesus, G. *et al.* Modulation of Hippocampal Plasticity and Cognitive Behavior by Short-term Blueberry Supplementation in Aged Rats. *Nutr. Neurosci.* **7**, 309–316 (2004).
89. Beltz, B. S., Tlusty, M. F., Benton, J. L. & Sandeman, D. C. Omega-3 fatty acids upregulate adult neurogenesis. *Neurosci. Lett.* **415**, 154–158 (2007).
90. Valente, T. *et al.* A Diet Enriched in Polyphenols and Polyunsaturated Fatty Acids, LMN Diet, Induces Neurogenesis in the Subventricular Zone and Hippocampus of Adult Mouse Brain. *J. Alzheimers Dis.* **18**, 849–865 (2009).
91. Dyall, S. C., Michael, G. J. & Michael-Titus, A. T. Omega-3 fatty acids reverse age-related decreases in nuclear receptors and increase neurogenesis in old rats. *J. Neurosci. Res.* **88**, 2091–2102 (2010).
92. Dong, S. *et al.* Curcumin Enhances Neurogenesis and Cognition in Aged Rats: Implications for Transcriptional Interactions Related to Growth and Synaptic Plasticity. *PLoS One* **7**, e31211 (2012).
93. Olson, A. K., Eadie, B. D., Ernst, C. & Christie, B. R. Environmental enrichment and voluntary exercise massively increase neurogenesis in the adult hippocampus via dissociable pathways. *Hippocampus* **16**, 250–260 (2006).
94. Alonso, M. *et al.* Olfactory Discrimination Learning Increases the Survival of Adult-Born Neurons in the Olfactory Bulb. *J. Neurosci.* **26**, 10508–10513 (2006).
95. Martončíková, M., Lievajová, K., Orendáčová, J., Blaško, J. & Račková, E. Odor enrichment influences neurogenesis in the rostral migratory stream of young rats. *Acta Histochem.* **113**, 326–332 (2011).
96. Rey, N. L., Sacquet, J., Veyrac, A., Jourdan, F. & Didier, A. Behavioral and cellular markers of olfactory aging and their response to enrichment. *Neurobiol. Aging* **33**, 626.e9–626.e23 (2012).
97. Bonzano, S., Bovetti, S., Fasolo, A., Peretto, P. & De Marchis, S. Odour enrichment increases adult-born dopaminergic neurons in the mouse olfactory bulb. *Eur. J. Neurosci.* **40**, 3450–3457 (2014).
98. Brown, J. *et al.* Enriched environment and physical activity stimulate hippocampal but not olfactory bulb neurogenesis. *Eur. J. Neurosci.* **17**, 2042–6 (2003).
99. Wojtowicz, J. M., Askew, M. L. & Winocur, G. The effects of running and of inhibiting adult neurogenesis on learning and memory in rats. *Eur J Neurosci* **27**, 1494–1502 (2008).
100. Creer, D. J., Romberg, C., Saksida, L. M., van Praag, H. & Bussey, T. J. Running enhances spatial pattern separation in mice. *Proc Natl Acad Sci U S A* **107**, 2367–2372

- (2010).
101. Sahay, A. *et al.* Increasing adult hippocampal neurogenesis is sufficient to improve pattern separation. *Nature* **472**, 466–470 (2011).
 102. Leasure, J. L. & Nixon, K. Exercise neuroprotection in a rat model of binge alcohol consumption. *Alcohol. Clin. Exp. Res.* **34**, 404–414 (2010).
 103. van Praag, H., Shubert, T., Zhao, C. & Gage, F. H. Exercise enhances learning and hippocampal neurogenesis in aged mice. *J Neurosci* **25**, 8680–8685 (2005).
 104. Hutton, C. P. *et al.* Synergistic effects of diet and exercise on hippocampal function in chronically stressed mice. *Neuroscience* **308**, 180–193 (2015).
 105. Veena, J. *et al.* Enriched environment restores hippocampal cell proliferation and ameliorates cognitive deficits in chronically stressed rats. *J. Neurosci. Res.* **87**, 831–843 (2009).
 106. Akers, K., Martinez-Canabal, a & Restivo, L. Hippocampal Neurogenesis Regulates Forgetting During Adulthood and Infancy. *Sci. (New York, N.Y.)cience* **598**, 598–602 (2014).
 107. Pereira, A. C. *et al.* An in vivo correlate of exercise-induced neurogenesis in the adult dentate gyrus. *Proc. Natl. Acad. Sci. U. S. A.* **104**, 5638–43 (2007).
 108. Manganas, L. N. *et al.* Magnetic resonance spectroscopy identifies neural progenitor cells in the live human brain. *Science (80-.)*. **318**, 980–985 (2007).
 109. Rueger, M. A. *et al.* Noninvasive Imaging of Endogenous Neural Stem Cell Mobilization In Vivo Using Positron Emission Tomography. *J. Neurosci.* **30**, 6454–6460 (2010).
 110. Couillard-Despres, S., Vreys, R., Aigner, L. & Van der Linden, A. In Vivo Monitoring of Adult Neurogenesis in Health and Disease. *Front. Neurosci.* **5**, (2011).
 111. Becker, S., MacQueen, G. & Wojtowicz, J. M. Computational modeling and empirical studies of hippocampal neurogenesis-dependent memory: Effects of interference, stress and depression. *Brain Res.* **1299**, 45–54 (2009).
 112. Shelton, D. J. & Kirwan, C. B. A possible negative influence of depression on the ability to overcome memory interference. *Behav Brain Res* **256**, 20–26 (2013).
 113. Déry, N. *et al.* Adult hippocampal neurogenesis reduces memory interference in humans: opposing effects of aerobic exercise and depression. *Front. Neurosci.* **7**, 66 (2013).
 114. Goldstein, A., Déry, N., Pilgrim, M., Ioan, M. & Becker, S. Stress and binge drinking: A toxic combination for the teenage brain. *Neuropsychologia* (2016). doi:10.1016/j.neuropsychologia.2016.07.035
 115. McNaughton, B. L. & Morris, R. G. M. Hippocampal synaptic enhancement and information storage within a distributed memory system. *Trends Neurosci.* **10**, 408–415 (1987).
 116. Treves, A. & Rolls, E. T. Computational constraints suggest the need for two distinct input systems to the hippocampal CA3 network. *Hippocampus* **2**, 189–200 (1992).
 117. O'Reilly, R. C. & McClelland, J. L. Hippocampal Conjunctive Encoding, Storage, and Recall: Avoiding a Tradeoff. *Hippocampus* **4**, 661–682 (1994).
 118. Faghihi, F. & Moustafa, A. A. A computational model of pattern separation efficiency in the dentate gyrus with implications in schizophrenia. *Front. Syst. Neurosci.* **09**, 42

- (2015).
119. Jung, M. W. & McNaughton, B. L. Spatial selectivity of unit activity in the hippocampal granular layer. *Hippocampus* **3**, 165–182 (1993).
 120. McClelland, J. L. & Goddard, N. H. Considerations Arising From Learning Systems Perspective on Hippocampus. *Hippocampus* **6**, 654–665 (1996).
 121. Becker, S. A computational principle for hippocampal learning and neurogenesis. *Hippocampus* **15**, 722–38 (2005).
 122. Myers, C. E. & Scharfman, H. E. A role for hilar cells in pattern separation in the dentate gyrus: a computational approach. *Hippocampus* **19**, 321–337 (2009).
 123. Leutgeb, J. K., Leutgeb, S., Moser, M.-B. & Moser, E. I. Pattern separation in the dentate gyrus and CA3 of the hippocampus. *Science (80-.)*. **315**, 961–966 (2007).
 124. Chawla, M. K. *et al.* Sparse, environmentally selective expression of Arc RNA in the upper blade of the rodent fascia dentata by brief spatial experience. *Hippocampus* **15**, 579–586 (2005).
 125. Satvat, E., Schmidt, B., Argraves, M., Marrone, D. F. & Markus, E. J. Changes in task demands alter the pattern of zif268 expression in the dentate gyrus. *J. Neurosci.* **31**, 7163–7167 (2011).
 126. Deng, W., Mayford, M. & Gage, F. H. Selection of distinct populations of dentate granule cells in response to inputs as a mechanism for pattern separation in mice. *Elife* **2013**, 1–21 (2013).
 127. Bakker, a., Kirwan, C. B., Miller, M. & Stark, C. E. L. Pattern Separation in the Human Hippocampal CA3 and Dentate Gyrus. *Science (80-.)*. **319**, 1640–1642 (2008).
 128. Marrone, D. F., Adams, A. A. & Satvat, E. Increased pattern separation in the aged fascia dentata. *Neurobiol. Aging* **32**, 2317.e23–2317.e32 (2011).
 129. Schmidt, B., Marrone, D. F. & Markus, E. J. Disambiguating the similar: The dentate gyrus and pattern separation. *Behav. Brain Res.* **226**, 56–65 (2012).
 130. Alme, C. B. *et al.* Hippocampal granule cells opt for early retirement. *Hippocampus* **20**, 1109–1123 (2010).
 131. Yassa, M. A. & Stark, C. E. L. Pattern separation in the hippocampus. *Trends Neurosci.* **34**, 515–525 (2011).
 132. O'Reilly, R. C. & Rudy, J. W. Conjunctive Representations in Learning and Memory: Principles of Cortical and Hippocampal Function. *Psychol. Rev.* **108**, 311–345 (2001).
 133. Finnegan, R. & Becker, S. Neurogenesis paradoxically decreases both pattern separation and memory interference. *Front. Syst. Neurosci.* **136**, (2015).
 134. Sahay, A., Wilson, D. A. & Hen, R. Perspective: Point/Counterpoint Pattern Separation: A Common Function for New Neurons in Hippocampus and Olfactory Bulb. *Neuron* **70**, 582–588 (2011).
 135. Ikrar, T. *et al.* Adult neurogenesis modifies excitability of the dentate gyrus. *Front. Neural Circuits* **7**, (2013).
 136. McAvoy, K., Besnard, A. & Sahay, A. Adult hippocampal neurogenesis and pattern separation in DG: a role for feedback inhibition in modulating sparseness to govern population-based coding. *Front. Syst. Neurosci.* **9**, (2015).
 137. Meltzer, L. A., Yabaluri, R. & Deisseroth, K. A role for circuit homeostasis in adult

- neurogenesis. *Trends Neurosci.* **28**, 653–660 (2005).
138. Aimone, J. B., Wiles, J. & Gage, F. H. Potential role for adult neurogenesis in the encoding of time in new memories. *Nat. Neurosci.* **9**, 723–727 (2006).
 139. Aimone, J. B., Wiles, J. & Gage, F. H. Computational Influence of Adult Neurogenesis on Memory Encoding. *Neuron* **61**, 187–202 (2009).
 140. Aimone, J. B., Deng, W. & Gage, F. H. Adult neurogenesis: Integrating theories and separating functions. *Trends Cogn. Sci.* **14**, 325–337 (2010).
 141. Rangel, L. M. *et al.* Temporally selective contextual encoding in the dentate gyrus of the hippocampus. *Nat. Commun.* **5**, (2014).
 142. Aimone, J. B., Deng, W. & Gage, F. H. Resolving New Memories: A Critical Look at the Dentate Gyrus, Adult Neurogenesis, and Pattern Separation. *Neuron* **70**, 589–596 (2011).
 143. Aimone, J. B. *et al.* Regulation and function of adult neurogenesis: from genes to cognition. *Physiol. Rev.* **94**, 991–1026 (2014).
 144. Johnston, S. T., Shtrahman, M., Parylak, S., Gonçalves, J. T. & Gage, F. H. Paradox of pattern separation and adult neurogenesis: A dual role for new neurons balancing memory resolution and robustness. *Neurobiol. Learn. Mem.* **129**, 60–68 (2016).
 145. Chambers, R. A., Potenza, M. N., Hoffman, R. E. & Miranker, W. Simulated apoptosis/neurogenesis regulates learning and memory capabilities of adaptive neural networks. *Neuropsychopharmacology* **29**, 747–758 (2004).
 146. Deisseroth, K. *et al.* Excitation-neurogenesis coupling in adult neural stem/progenitor cells. *Neuron* **42**, 535–552 (2004).
 147. Weisz, V. I. & Argibay, P. F. Neurogenesis interferes with the retrieval of remote memories: Forgetting in neurocomputational terms. *Cognition* **125**, 13–25 (2012).
 148. Frankland, P. W., Köhler, S. & Josselyn, S. A. Hippocampal neurogenesis and forgetting. *Trends Neurosci.* **36**, 497–503 (2013).
 149. Josselyn, S. A. & Frankland, P. W. Infantile amnesia: a neurogenic hypothesis. *Learn. Mem.* **19**, 423–433 (2012).
 150. Frankland, P. W., O'Brien, C., Ohno, M., Kirkwood, A. & Silva, A. J. Alpha-CaMKII-dependent plasticity in the cortex is required for permanent memory. *Nature* **411**, 309–313 (2001).
 151. Déry, N., Goldstein, A. & Becker, S. A Role for Adult Hippocampal Neurogenesis at Multiple Time Scales : A Study of Recent and Remote Memory in Humans. *Behav. Neurosci.* **129**, 435–449 (2015).
 152. Rumelhart, D. ~E. & Zipser, D. in *Parallel Distrib. Process. Explor. Microstruct. Cogn.* (Rumelhart, J. ~L. M. D. ~E. & the PDP research group) **I**, (Bradford Books, 1986).
 153. Grossberg, S. Adaptive pattern classification and universal recoding, {I}: Parallel development and coding of neural feature detectors. *Biol. Cybern.* **23**, 121–134 (1976).
 154. Grossberg, S. Competitive Learning: From Interactive Activation to Adaptive Resonance. *Cogn. Sci.* **11**, 23–63 (1987).
 155. Bergami, M. *et al.* A Critical Period for Experience-Dependent Remodeling of Adult-Born Neuron Connectivity. *Neuron* **85**, 710–717 (2015).

156. Dieni, C. V., Nietz, A. K., Panichi, R., Wadiche, J. I. & Overstreet-Wadiche, L. Distinct Determinants of Sparse Activation during Granule Cell Maturation. *J. Neurosci.* **33**, 19131–19142 (2013).
157. Grossberg, S. Adaptive Pattern Classification and Universal Recoding: {II}. Feedback, Expectation, Olfaction, Illusions. *Biol. Cybern.* **23**, 187–202 (1976).
158. Nolan, C. R., Wyeth, G., Milford, M. & Wiles, J. The race to learn: Spike timing and STDP can coordinate learning and recall in CA3. *Hippocampus* **21**, 647–660 (2011).
159. Tronel, S. *et al.* Adult-born dentate neurons are recruited in both spatial memory encoding and retrieval. *Hippocampus* **25**, 1472–1479 (2015).
160. Suárez-Pereira, I. & Carrión, Á. M. Updating stored memory requires adult hippocampal neurogenesis. *Sci. Rep.* **5**, 13993 (2015).
161. Jacobs, B. L., van Praag, H. & Gage, F. H. Adult brain neurogenesis and psychiatry: a novel theory of depression. *Mol. Psychiatry* **5**, 262–269 (2000).
162. Gould, E., Tanapat, P., McEwen, B. S., Flügge, G. & Fuchs, E. Proliferation of granule cell precursors in the dentate gyrus of adult monkeys is diminished by stress. *Proc. Natl. Acad. Sci. U. S. A.* **95**, 3168–3171 (1998).
163. Malberg, J. E., Eisch, A. J., Nestler, E. J. & Duman, R. S. Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. *J. Neurosci* **20**, 9104–9110 (2000).
164. Kodama, M., Fujioka, T. & Duman, R. S. Chronic olanzapine or fluoxetine administration increases cell proliferation in hippocampus and prefrontal cortex of adult rat. *Biol. Psychiatry* **56**, 570–580 (2004).
165. Scott, B. W., Wojtowicz, J. M. & Burnham, W. M. Neurogenesis in the dentate gyrus of the rat following electroconvulsive shock seizures. *Exp. Neurol.* **165**, 231–6 (2000).
166. Madsen, T. M. *et al.* Increased neurogenesis in a model of electroconvulsive therapy. *Biol Psychiatry* **47**, 1043–1049 (2000).
167. Lyons, D. M. *et al.* Stress coping stimulates hippocampal neurogenesis in adult monkeys. *Proc. Natl. Acad. Sci. U. S. A.* **107**, 14823–7 (2010).
168. Warner-Schmidt, J. L., Madsen, T. M. & Duman, R. S. Electroconvulsive seizure restores neurogenesis and hippocampus-dependent fear memory after disruption by irradiation. *Eur. J. Neurosci.* **27**, 1485–1493 (2008).
169. Deng, W. & Gage, F. H. The effect of immature adult-born dentate granule cells on hyponeophagial behavior is related to their roles in learning and memory. *Front. Syst. Neurosci.* **9**, (2015).
170. Schloesser, R. J., Manji, H. K. & Martinowich, K. Suppression of adult neurogenesis leads to an increased hypothalamo-pituitary-adrenal axis response. *Neuroreport* **20**, 553–7 (2009).
171. Snyder, J. S., Soumier, A., Brewer, M., Pickel, J. & Cameron, H. A. Adult hippocampal neurogenesis buffers stress responses and depressive behaviour. *Nature* **476**, 458–61 (2011).
172. Santarelli, L. *et al.* Requirement of hippocampal neurogenesis for the behavioral effects of antidepressants. *Science (80-.)*. **301**, 805–809 (2003).
173. Perera, T. D. *et al.* Necessity of hippocampal neurogenesis for the therapeutic action of antidepressants in adult nonhuman primates. *PLoS One* **6**, e17600 (2011).

174. Besnard, A. & Sahay, A. Adult Hippocampal Neurogenesis, Fear Generalization, and Stress. *Neuropsychopharmacology* **41**, 24–44 (2016).
175. Sahay, A. & Hen, R. Adult hippocampal neurogenesis in depression. *Nat. Neurosci.* **10**, 1110–1115 (2007).
176. Becker, S. & Wojtowicz, J. M. A model of hippocampal neurogenesis in memory and mood disorders. *Trends Cogn. Sci.* **11**, 70–76 (2007).
177. Ekdahl, C. T., Claassen, J.-H., Bonde, S., Kokaia, Z. & Lindvall, O. Inflammation is detrimental for neurogenesis in adult brain. *Proc. Natl. Acad. Sci.* **100**, 13632–13637 (2003).
178. Daulatzai, M. A. Neurotoxic saboteurs: Straws that break the Hippo's (Hippocampus) back drive cognitive impairment and Alzheimer's disease. *Neurotox. Res.* **24**, 407–459 (2013).
179. Solano Fonseca, R. *et al.* Neurogenic Niche Microglia Undergo Positional Remodeling and Progressive Activation Contributing to Age-Associated Reductions in Neurogenesis. *Stem Cells Dev.* **25**, 542–555 (2016).
180. Kempermann, G., Kuhan, H. G. & Gage, F. H. Experience-induced neurogenesis in the senescent dentate gyrus. *J. Neurosci.* **18**, 3206–3212 (1998).
181. Kempermann, G., Gast, D. & Gage, F. H. Neuroplasticity in old age: Sustained fivefold induction of hippocampal neurogenesis by long-term environmental enrichment. *Ann. Neurol.* **52**, 135–143 (2002).
182. Cotman, C. W., Berchtold, N. C. & Christie, L.-A. Exercise builds brain health: key roles of growth factor cascades and inflammation. *Trends Neurosci.* **30**, 464–472 (2007).
183. Bernadeta Michalski, Maria M. Corrada, Claudia H. Kawas, M. F. Brain-derived neurotrophic factor and TrkB expression in the 'oldest-old,' the 90p Study: correlation with cognitive status and levels of soluble amyloid-beta. *Neurobiol. Aging* **36**, 3130–3139 (2015).
184. Siette, J. *et al.* Age-specific effects of voluntary exercise on memory and the older brain. *Biol. Psychiatry* **73**, 435–442 (2013).
185. Fabel, K. *et al.* Additive effects of physical exercise and environmental enrichment on adult hippocampal neurogenesis in mice. *Front Neurosci* **3**, 50 (2009).
186. Kempermann, G. *et al.* Why and how physical activity promotes experience-induced brain plasticity. *Front. Neurosci.* **4**, 189 (2010).
187. Fahnstock, M. *et al.* BDNF increases with behavioral enrichment and an antioxidant diet in the aged dog. *Neurobiol. Aging* **33**, 546–554 (2012).

Figure captions

Figure 1. Schematic models of the hippocampus, using sparse coding (A) versus neurogenesis and sparse connectivity (B, C) to overcome interference. Model input is a distributed pattern of activation

across the entorhinal cortex pyramidal cells (PC; activated cells shown in pink). This input generates a sparse pattern of activation across the mature dentate granule cells (GCs; active cells shown in blue). PCs are densely interconnected to mature GCs, which are themselves interconnected via inhibitory interneurons (IN, shown in yellow). In B) and C), the model with neurogenesis also includes immature GCs (active cells shown in green); entorhinal PCs are sparsely connected to the immature GCs. The model shown in B) and C) is presented with two different input patterns and in response, generates identical sparse activation patterns in mature GCs, and overlapping but distinct distributed activation in immature GCs. While the two input patterns overlap by 40%, the two patterns of activation in the immature GC overlap by 50%, hence a decrease in pattern separation. The model is nonetheless capable of maintaining distinct neural codes for the two similar inputs, in spite of high plasticity in the immature cell population, due to the sparse connectivity of the immature GCs.

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